



Antimicrobial Effect of *Cyperus rotundus* on Multiple Drug Resistant *Pseudomonas aeruginosa* Strains

Mohammad Dadook¹, Sedigheh Mehrabian¹, Saeed Irian^{2*}

¹ Faculty of Biological Sciences, Islamic Azad University, Tehran North Branch, Tehran, Iran.

² Department of Cell and Molecular Biology, Faculty of Biological Sciences, Kharazmi University, Tehran, Iran.

ARTICLE INFO

Article type:
Original Article

Article history:

Received: 12 Dec 2015
Revised: 22 Jan 2016
Accepted: 14 May 2016
Published: 15 Aug 2016

Keywords:

Cyperus rotundus,
Antibacterial,
Pseudomonas aeruginosa, Medicinal
plants

ABSTRACT

Background: Medicinal use of plants dates long back in the history of the human beings and has recently gained a great deal of importance in treating different diseases. Due to the importance of *Pseudomonas aeruginosa* strains as agents responsible for common secondary infections and their resistance to antibiotics and disinfectants, the present study aimed at investigating the antimicrobial effects of the plant *Cyperus rotundus* on multiple drug resistant (MDR) *P. aeruginosa* strains.

Methods: Ethanolic extracts of *Cyperus rotundus* tuber were prepared by maceration. The antimicrobial effect of these extracts on *P. aeruginosa* isolates (ie., sensitive to antibiotics and multiple drug resistant strains), isolated from clinical and soil samples, was determined by both disk diffusion and broth microdilution methods..

Results: It was revealed that ethanolic extract concentrations of *Cyperus rotundus* tuber higher than 0.1 mg/ml suppresses the growth of all antibiotic sensitive and resistant *P. aeruginosa* strains which were used in this study.

Conclusion: It is concluded that *Cyperus rotundus* possesses antimicrobial properties and thus can be used in treating multidrug resistant *P. aeruginosa*-induced wounds and infections.

- **Please cite this paper as:** Dadook M, Mehrabian S, Irian S. Antimicrobial Effect of *Cyperus rotundus* on Multiple Drug Resistant *Pseudomonas aeruginosa* Strains. *J Med Bacteriol.* 2016; **5** (1, 2): pp.15-20.

Introduction

Pseudomonas aeruginosa is an aerobic, opportunistic, Gram negative bacterium with lipopolysaccharide, polar flagella and pili (1, 2). The bacterium is distributed widely in nature and acts as an infecting agent in patients inflicted with immune deficiency, neutropenic, burns and catheter. It is also the most common agent in infecting the respiratory, urogenital, digestive and the central nervous systems, as well as bone and cartilage tissues, skin and soft tissue, bacteremia, septicemia and eye and ear infections (3, 4, 5).

Treatment of *Pseudomonas aeruginosa* infected patients, in particular those infected with the antibiotic resistant strains, is not an easy task (6). Following inappropriate experimental treatments, sensitive microorganisms also turn resistant, a process requiring the induction of the antibiotic inactivating enzymes or mutations in genes encoding outer membrane porins, or plasmid transfer. Beta lactamase production is one of the most important mechanisms of bacterial resistance, and the number of beta-lactamase producing bacteria are on the rise due to mutations (7, 8).

Plants have served as medicine since the ancient times (9). Similar to antibiotics, the antimicrobial effects of medicinal plants have gained a great deal of attention (10, 11). Use of medicinal plants in the past several decades has increasingly turned popular such that a great number of the available medicines are of plant origin (12, 13), and medicinal plants are currently being used for the treatment of many diseases including fungal infections (14). The aim of the present study was to determine the contamination degree of tap waters with *P. aeruginosa*, *L. pneumophila*, and *E. coli* in all cities in Guilan province, Iran.

Cyperus rotundus, also known as nut grass, as a member of the Cyperaceae family is distributed in humid, marshy, warm and moderate environments including North Iran with a local name of "Teplagh". It is a perennial plant with 20-40 cm height, long roots with rhizomes and a black shelled tuber with a white and odorous interior. Leaves are numerous, small, linear, dark green, and

spikelets are reddish brown (15, 16, 17). The rhizome contains essential oil, saponins, vitamin C, polyphenol and flavonol glycoside (18, 19). *Cyperus rotundus* has had medicinal use in gastrointestinal bloating, stomach burning, stomach ulcers, menstrual periods, kidney problems, headaches and liquid mouthwash (20, 21, 22).

Due to the increasing importance of medicinal plants in herbal medicine and their low rate of side effects, and the significant hospital infections caused by *Pseudomonas aeruginosa* and its resistance to most antibiotics, this study aimed to investigate the antimicrobial effects of the ethanolic extract of *Cyperus rotundus* on both sensitive and multidrug resistant *Pseudomonas aeruginosa* strains.

Material and method

Isolation and purification of Pathogenic microorganisms

To isolate and purify *Pseudomonas aeruginosa* strains, including; 40 clinical samples from patient wounds, hospitalized at Motahari Hospital, Tehran, Iran. Also 50 soil samples from Eznova-Behnemir region Mazandaran-Iran were prepared in sterile tubes and transferred to the laboratory under sterile conditions. Serial dilutions of 10^{-1} - 10^{-10} were prepared from samples and plated on citrimide medium in duplicates. Plates were incubated at 37 °C for 48 h and bacterial colonies were subjected to both macroscopic and microscopic examinations. *Pseudomonas aeruginosa* strain identification was performed by Gram staining, oxidase, catalase, pigmentation, OF, citrate, urease, arginine dehydrolase, lysine and ornithine decarboxylase tests, interaction in TSI medium, and growth at 42 °C (23).

Cyperus rotundus tuber extract preparation

Cyperus rotundus tubers were collected from the agricultural lands of Aznava-Behnamir in Mazandaran province of Iran. Extracts were

prepared by maceration or wetting by ethanol solution. Tubers were dried in a closed environment with ventilation, and then powdered. A total of 200 grams of the powder was added to 100 ml of ethanol and incubated for 48 h, and then filtered through a sterile paper filter. This filtered extract was then dried by incubation at 40 °C.

Strain sensitivity to antibiotics

Initially, microbial suspensions of the bacteria with a turbidity of half a McFarlen (1.5×10^8) were prepared according to CLSI (The Clinical and Laboratory Standards Institute). Samples of microbial suspensions were grown on Muller Hinton plates using a sterile swap. Bacterial sensitivity to antibiotics Amikacin (30 µg), Ciprofloxacin (5 µg), Piperacillin (100 µg), Ceftriaxone (30 µg), Ceftazidime (30 µg), Cefepime (30 µg), Imipenem (10 µg), Meropenem (10 µg) was tested using antibiotic discs (Padtan Teb, Iran) (Kirby bauer) (24). Following incubation at 37 °C for 18-24 h, the diameter of the zone of inhibition around the disc was measured. Bacterial strain sensitivity or resistance to antibiotics was determined according to the standard table of CLSI.

Antimicrobial activity of the tuber extract

Strain sensitivity was determined by both disk diffusion and broth microdilution methods. Initially, suspensions of 0.5 McFarlen (1.5×10^8 CFU/ml) were prepared (25).

In the disc diffusion method, using a sterile swap, suspensions were grown on Muller Hinton agar medium. Sterile blank discs were placed in plates containing different concentrations of the tuber extract and then placed on agar plates containing microorganisms. These plates were incubated at 37 °C for 24 h, and then subjected to measurements of the zone of growth inhibition (17).

In the broth microdilution method, tubes containing Muller Hinton broth medium containing 0.1-20 mg/ml of the tuber extract to a total volume of 1 ml were prepared and inoculated

with 100 µl of 0.5 McFarlen suspension (26). A solution of 0.1 mg/ml of the extract was also included as negative control. MIC values were determined after 24 h of incubation at 37 °C (27).

Results

In total a collection of 84 *Pseudomonas aeruginosa* strains were isolated from both clinical (50 strains) and soil samples (34 strains) and identified by biochemical tests. Of these, 21 strains were multidrug resistant (19 from clinical and 2 from soil samples) (Table 1).

Table 1. Results of the antibiogram test in multiple drug resistant *Pseudomonas aeruginosa* strains.

Antibiotic	# of Resistant Strains	# of Resistant Clinical Strains	# of Resistant Soil Strains
Total number of resistant strains	21	19	2
Amikacin	15	15	0
Ciprofloxacin	10	10	0
Piperacillin	13	13	0
Piperacillin	21	19	2
Ceftazidime	20	19	1
Cefepime	20	19	1
Imipenem	21	19	2
Meropenem	21	19	2

Strain sensitivity to *Cyperus rotundus* tuber extract

In the blank disc method, zone of growth inhibition appeared at a minimum inhibitory concentration of 25 mg/ml for all bacterial strains (Table 2).

Table 2. Blank disc results on the inhibitory effect of *Cyperus rotundus* extract on *Pseudomonas aeruginosa*.

Microorganism	Diameter of zone of growth inhibition (mm) at 25 mg/ml of the extract
Antibiotic sensitive <i>Pseudomonas aeruginosa</i> strains	15
Multiple drug resistant <i>Pseudomonas aeruginosa</i> strains	15

In the broth microdilution method, the MIC value for all bacterial strains was at a concentration of 0.1 mg/ml (Table 3).

Table 3. MIC values of the ethanolic extract of *Cyperus rotundus* on the growth of *Pseudomonas aeruginosa*.

Microorganism	MIC value (mg/ml)
Antibiotic sensitive <i>Pseudomonas aeruginosa</i> strains	0.1
Multiple drug resistant <i>Pseudomonas aeruginosa</i> strains	0.1

Discussion

In the present study, 84 *Pseudomonas aeruginosa* strains resistant and sensitive to antibiotics, prepared from both clinical and soil samples, were isolated using citrimide agar medium, and identified based on phenotypic and biochemical analyses. These bacterial strains were then treated with the ethanolic extract of *Cyperus rotundus* tuber. It was revealed that the tuber extract possesses an antimicrobial effect, such that the antibiotic resistant strains did not show any resistance to the extract, and the MIC value for all the strains was determined to be 0.1 mg/ml.

Cyperus rotundus tuber extract has been shown to possess antimicrobial activity (28). While, its inhibitory effect against *Streptococcus pyogenes* growth was demonstrated by Mehta et al. (29), the whole plant extract has been shown to be ineffective against strains of *E. coli*, *Pseudomonas aeruginosa* and *Salmonella typhi* (30). A MIC value of 12.5 mg/ml has indeed been determined for certain bacterial strains treated with the rhizomes oil (17), whereas using the disc diffusion method, zone of growth inhibition was revealed in *Pseudomonas aeruginosa* and *Staphylococcus aureus* treated with the ethanolic extract of the plant (31). In a separate study, using the agar disk diffusion method, the ethanolic extract of *Cyperus rotundus* revealed a zone of growth inhibition for *E. coli* and *C. albicans* (32).

Conclusion

In conclusion, *Cyperus rotundus* extract possesses an antimicrobial effect and inhibits the growth of resistant *Pseudomonas aeruginosa* and thus maybe considered as an effective agent in treating patients infected with MDR strains of *Pseudomonas aeruginosa*. Considering the increasing trend in the use of antibiotics along with the major clinical and public health problems associated with antibiotic resistance, plant based antimicrobial agents, such as those contained within *Cyperus rotundus* extract, will be of great use in both preventing and curing *Pseudomonas aeruginosa* related infections.

Conflict of interests

No conflict of interests is declared.

Financial disclosure

Authors declare no financial disclosures.

References

1. Driscoll JA, Brody SL, Kollef MH. The epidemiology, pathogenesis and treatment of *Pseudomonas aeruginosa* infections. *Drugs* 2007; **67**(3): 351-368.
2. Brooks GF, Butel JS, Morse SA. Jawet Melnick and Adelberg's Medical Microbiology. London: Lang Basic Science 2004; pp: 262-267.
3. Akhabue E, Synnestvedt M, Weiner M G, et al. Cefepime resistant *Pseudomonas aeruginosa*. *Emerg Infect Dis* 2011; **17**(6): 1037-43.
4. Aoki S, Hirakata Y, Kondoh A, et al. Virulence of metallo-beta-lactamase producing *Pseudomonas aeruginosa* In vitro. *Antimicrob Agents Chemother* 2004; **48** (5): 1876-1878.
5. Doring G, Pier GB. Vaccines and immunotherapy against *Pseudomonas aeruginosa*. *Vaccine* 2008; **26**: 1011-1024.

6. Maleknezhad P, Aligholi M, Moosavi S. Study of *Pseudomonas aeruginosa* resistance to penicillins, cephalosporins and aminoglycosides. *The Univ Med J* 1998; **56** (4): 23-29.
7. Church D, Elsayed S, Reid O, et al. Burn wound infections. *Clin Microbiol Rev* 2006; **19**: 403-34.
8. Japoni A, Alborzi A, Kalani M, et al. Susceptibility patterns and cross resistance of antibiotics against *Pseudomonas aeruginosa* isolated from burn patients in the south of Iran. *Burns* 2006; **32**(3): 343-7.
9. Cowan MM. Plant products as antimicrobial agents. *Clin Microbiol Rev* 1999; **12**(4): 564-82.
10. Alviano DS, Alviano CS. Plant extracts: search for new alternatives to treat microbial diseases. *Curr Pharm Biotechnol* 2009; **10**: 106-21.
11. Kumar P, Ansari SH, Ali J. Herbal remedies for the treatment of periodontal disease. *Recent Pat Drug Deliv Formul* 2009; **3**(3): 221-28.
12. Grag SC, Denger SL. Antifungal activity of the essential oil of *Myrtus communis* var. *microphylla*. *Herba Hungarica* 1988; **27**(2-3): 123-124.
13. Hines T, Hill T. Encyclopedia of medicinal plants. 15th ed. London: *Dorsley Kindersley* 1996; pp: 23-60.
14. Falahati M, Tabrizi ON, Jahanian F. Antidermatophyte activity of *Eucalyptus camadulensis* in comparison with griseofulvin. *Iranian J Pharm & Ther* 2005; **4**: 80-83.
15. Burkill H. The useful plants of west tropical Africa. Vol. 1. 2nd ed. Kew (UK); Royal Botanic Gardens 1985.
16. Gunasekera TGLG, Fernando DNS. Agricultural importance, biology, control and utilization *Cyperus rotundus*. *The Planter, Kuala Lumpur* 1994; **70**: 537-544.
17. Eltayeib AA, Ismaeel HU. Extraction of *Cyperus rotundus* rhizomes oil, identification of chemical constituents and evaluation of antimicrobial activity of the oil in North kordofan state. *International Journal of Advanced Research in Chemical Science (IJARCS)* 2014; **1**(9): 18-29.
18. Adeniyi TA, Adeonipekun PA, Omotayo AE. Investigating the phytochemicals and Antimicrobial properties of three sedge (cyperaceae) species. *International journal of tropical medicine* 2013; **8**(4): 92-98.
19. Sivapalan SR. Physico-chemical and phytochemical study of rhizome of *Cyperus rotundus* LINN. *International Journal of Pharmacology and Pharmaceutical Technology (IJPPT)* 2012; **1**(2): 42-46.
20. Raut NA, Gaikwad NJ. Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia* 2006; **77**(7-8): 585-588.
21. Uddin SJ, Mondal K, Shilpi JA, et al. Antidiarrhoeal activity of *Cyperus rotundus*. *Fitoterapia* 2006; **77**(2): 134-136.
22. Muthu K, Hema M, Nagaraj S, et al. In-vitro antibacterial potential, phytochemical characterization of *Cyperus rotundus* flower extract. *Int. j. appl. res. nat. prod* 2014; **4**(1): 6-8.
23. Forbes BA, Sahm DF, Weissfeld AS. *Bailey & Scott's Diagnostic Microbiol.* 12th ed. Missouri. Mosby 2007.
24. Washington JA, Anhatt JP. Antimicrobial susceptibility tests of aerobic and facultatively anaerobic bacteria. In: *Laboratory procedures in clinical microbiology.* 2nd ed. USA. Springer 1985.
25. Chessbrough M. *District laboratory practice in tropical countries.* United Kingdom. Cambridge Univ Press 2000.
26. Harris DC. *Quantitative chemical analysis,* Eighth Edition. W H Freeman and Co. New York NY 2010.

27. Sharma SK, Singh AP. Antimicrobial investigations on rhizomes of *Cyperus rotundus* Linn. *Der Pharmacia Lettre* 2011; **3**(3): 427-431.
28. Bisht A, Bisht GRS, Singh M, et al. Chemical composition and antimicrobial activity of essential oil of tubers of *Cyperus rotundus* Linn. collected from Dehradun (Uttarakhand). *IJRPBS* 2011; **2**(2): 661-665.
29. Mehta M, Bharmuche A, Bhatkala A. Investigation of the anti-microbial and anti-inflammatory effect of *Cyperus rotundus* on Tonsillitis. *IJCET* 2013; **27**: 135-138.
30. Parekh J, Chanda C. In-vitro antimicrobial activities of extracts of *Launaea procumbens* Roxb. (Labiatae), *Vitis vinifera* L. (Vitaceae) and *Cyperus rotundus* L. (Cyperaceae). *Afr J Biomed Res* 2006; **9**(2): 89-93.
31. Tasleem A, Mateen A, Abdul Waheed M, et al. Antimicrobial activity of some herbal drugs used in unani system of medicine. *Int j herb med* 2015; **2**(5): 27-30.
32. Kabbashi AS, Eldeen AS, Almagboul MAZ, et al. Antimicrobial activity and cytotoxicity of ethanolic extract of *Cyperus rotundus* L. *American Journal of Pharmacy and Pharmaceutical Sciences* 2015; **2**(1): 1-13.