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Molecular Characterization of Exotoxin Genes in *Staphylococcus aureus* Recovered From Hospitalized Patients

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ARTICLE INFO	ABSTRACT
Article type: Research Article	Background : Staphylococcus aureus is considered as a major cause of skin and soft tissue infections, arthritis, osteomyelitis, infective endocarditis, and pneumoniae though community or
Article history: Received: 17 Jun 2019 Revised: 26 Jul 2019 Accepted: 07 Aug 2019 Published: 05 Sep 2019	nosocomial transmission. In this study, attempts were made to investigate the distribution of some important exotoxin genes, including <i>hla</i> , <i>hlb</i> , <i>tsst</i> -1, <i>eta</i> , <i>etb</i> , and <i>etd</i> among methicillin-resistant <i>S. aureus</i> (MRSA) isolated from a hospital patients in Tabriz, Iran. <i>Methods</i> : In the present cross-sectional study, a total of 90 <i>S. aureus</i> were isolated from children who admitted to a hospital during six-month in 2017. Isolates were identified using biochemical tests and then, using PCR, the isolates were tested for the presence of, <i>hla</i> , <i>hlb</i> , <i>tsst</i> -1, <i>eta</i> , <i>etb</i> , and <i>etd</i>
Keywords: Exotoxins, Methicillin- resistant Staphylococcus aureus6, toxic shock syndrome toxin-1, Pediatrics, Staphylococcal exfoliative toxin.	genes. Results: It was found that 40% of the <i>S. aureus</i> were considered as MRSA strains by biochemical and molecular tests. The results of molecular detection of virulence determinants showed that <i>eta</i> , <i>hla</i> , <i>etb</i> , <i>tsst</i> -1, <i>hlb</i> and <i>etd</i> were detected in 86.1%, 80.5%, 30.5%, 27.7%, 22.2%, and 19.4% of isolates, respectively. Conclusion: Our findings clarify characterization of toxin production status of <i>S. aureus</i> isolates from patients in Iran. The current study showed that a majority of <i>S. aureus</i> isolates harbored <i>eta</i> and <i>hla</i> virulence gene.

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Introduction

Staphylococcus aureus has been considered as an important human pathogen causing a wide range of infections in both hospital and community settings (1). This ever-present opportunistic pathogen is among the most common causes of skin and soft tissue infections, arthritis, osteomyelitis, infective endocarditis, pneumoniae, as well as nosocomial infections, including various surgical infections and bacteremia (2, 3). As a matter of fact, neonates, infants and young children are more likely to be affected by *S. aureus* infections, notwithstanding whether these infections had their onset in the community or were acquired in the hospital (4).

A characteristic feature of pathogenic S. aureus strains is the expression of a broad range of adhesion proteins and virulence factors that play a significant role on every level of host-pathogen interactions during different phases of colonization and infection (5). These virulence factors allow the bacterium to escape the immune responses, attach to the host tissue, and cause severe damage to the host tissues (6, 7). Exotoxins such as hemolysins (Hla, Hlb, Hld, and panton-valentine leukocidin (PVL), Hlg), Exfoliative toxin A and B (*EtA* and *EtB*), Staphylococcal Enterotoxins (SEs), Toxic shock syndrome toxin-1 (TSST-1) are the most important virulence factors produced by this pathogen (8). However, it should be pointed out that most of these factors may not exist in all stains of S. aureus and their expression occurs in specified strains and under certain conditions. For instance, the S. aureus strains that produce PVL produce PVL are associated with communityacquired hemorrhagic and necrotizing pulmonary infections affecting previously healthy children and young adults (9, 10).

In addition to the high pathogenicity of *S. aureus*, the pathogen is well identified for its capacity to acquire resistance to several antibiotics. In the early of 1960s, methicillin-

resistant S. aureus (MRSA) strains have emerged as significant pathogens of nosocomialand community acquired- infections and then ultimately spread worldwide over the next several decades (11, 12). The mecA gene is part of the chromosome staphylococcal cassette mec (SCCmec) which is responsible resistance to methicillin and other beta-lactam antibiotics (13). Several researchers have indicated that MRSA strains have become not only resistant to betalactam agents, but they can also acquire additional resistance to other antibiotic classes, especially aminoglycosides (14-16). Moreover, the presence of resistance genes, such as *mecA*, may also affect toxin production among S. aureus strains (17).

Considering of the increasing spread of MRSA strains among adults and pediatrics population, the current study was conducted to detect the virulence determinants, including hla, hlb, eta, etb, etd, and tsst-1, among MRSA strains isolated from patients in Tabriz, Iran. Staphylococcus aureus has been considered as an important human pathogen causing a wide range of infections in both hospital and community settings (1). This ever-present opportunistic pathogen is among the most common causes of skin and soft tissue infections, arthritis, osteomyelitis, infective endocarditis, pneumoniae, as well as nosocomial infections, including various surgical infections and bacteremia (2, 3). As a matter of fact, neonates, infants and young children are more likely to be affected by S. aureus infections, notwithstanding whether these infections had their onset in the community or were acquired in the hospital (4).

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Materials and Methods

Bacterial Isolation and Identification

The present study 90 *S. aureus* isolates were obtained from /hospitalized patients with clinical signs and symptoms of colonization such as increased fever and white and blood cell counts admitted to hospital, during 6-month Tabriz, Iran. Standard conventional biochemical tests were carried out on colonies from primary cultures in order to identification of the bacterial isolates. Accordingly, all suspected colonies of *S. aureus* were examined using haemolysis, gram staining, catalase, oxidase, DNase, coagulase and mannitol fermentation tests. The confirmed isolates were stored at -70 °C in trypticase soy broth (TSB) (Merck Co., Germany) containing 20% glycerol for later analysis.

Detection of MRSA Strains

Phenotypic detection of MRSA isolates was performed using disk diffusion method according to the Clinical and Laboratory Standards Institute (CLSI) 2017 guidelines with a 30 µg cefoxitin disc (Mast Co., UK) on Mueller–Hinton agar followed by incubation at 30° C for 24 hr. The isolates that exhibited inhibition zone size ≤ 19 mm were considered as MRSA strains.

Molecular Assay for Detection of Virulence Genes

Total DNA of the isolates was extracted via High Pure PCR Template Preparation Kit (Thermo Fisher Co., USA) according to the manufacturer's instructions. The presence of the virulence determinants including *hla*, *hlb*, *eta*, *etb*, etd, and tsst-1 were detected by separate PCR reactions, using specific primers (Table 1). All PCR reactions were performed in a final volume of 25 µL containing the 12.5 µl of master mix (Cinnaclon, Tehran, Iran), 8.5 µl of distilled water, 10 pmol of each primer, and 1 µL DNA template (3 μ g/ μ L). The thermal cycling condition of the mixture included an initial denaturation step at 95 °C for 5 minutes followed by 35 cycles of amplification comprising of three steps: denaturation at 94 °C for 1 min; 30 s at different annealing temperatures for each primer set (Table 1), and with primer extension at 72°C for 45 s. The final extension step was done at 72°C for 5 minutes. The PCR products were analysed using Tris/Borate/EDTA (TBE) buffered agarose gel (1.5%) electrophoresis at 120 V for 45 minutes.

Statistical Analyses

Categorical variables were calculated by using SPSS 22.0 statistical software (IBM Corp., Armonk, USA). A statistically significant difference was considered if P value was <0.05.

Results

Patients and Bacterial Identification

In this study, 90 clinical isolates of *S. aureus* were collected from adults (18-85 year) and paediatrics that was under 18 years of age during a six-month period. Of these, 17 isolates were

collected (43.5%) from outpatient or those referred to the emergency department, with the remainder from different wards, including infection (23%), internal (20.5%), and pediatric units (13%). In total, 19 (52.77%) clinical isolates of *S. aureus* were recovered from blood, and the remaining were obtained from urine (16%), wound (10.7%), tracheal (10.3%), synovial (5.1%), bronchial (2.6%), and abscess (2.6%). Among 90 *S. aureus* isolates, 36 (40%) were recognized as MRSA strains by phenotypic method. In addition, the distributions of *mecA* gene among MRSA isolates were 100%.

Presence of Virulence Determinants

The results of molecular detection of virulence determinants showed that *eta*, *hla*, *etb*, *tsst*-1, *hlb* and *etd* were detected in 86.1%, 80.5%, 30.5%, 27.7%, 22.2%, and 19.4% of isolates, respectively. Table 2 illustrates the distribution of the virulence determinants among *S. aureus* isolates from patients in this study.

Discussion

The production of exotoxins in clinical isolates of S. aureus is the most important factor in the development of severe infection and allows for immune evasion (18). Specific staphylococcal exotoxins seem to be responsible for individual syndromes caused by the bacterium. For example, scalded skin syndrome (SSSS) has been associated with ETA and ETB and toxic shock syndrome with TSST-1 (19). Hence, in order to improve surveillance, control and treatment of S. aureus infections among pediatric patients, it is essential to understand the biofilm formation that lead to chronic infections (20). In the current study, we focused on evaluation of virulence gene profiles of clinical isolates of MRSA isolated from patients, which may affect the severity of infection among patients.

Using of phenotypic and molecular assays a relatively high frequency of MRSA strains was identified and is in accordance with the findings reported by Esmaeili Benvidi et al. among paediatric patients in Tehran (21). However, the higher prevalence of MRSA infections has been previously reported in Iranian population (22, 23). These variations may detected be attributed to the efficiency of screening methods for MRSA identification. Moreover, the variations in the results can be clarified by the fact that the other researches were conducted on all age groups or healthy carrier persons, especially in pediatrics patients.

In the present study, it was found that a majority of the MRSA isolates, recovered from patients, harbored eta and hla virulence gene particularly all of pediatrics patients in our study detected *hla* gene. However, in a cross-sectional study, Esmaeili Benvidi et al. described that none of the investigated MRSA isolates from pediatric patients harbored eta gene in Tehran (21). In addition, the low detection frequency of eta (11.3%) were previously reported in a study conducted by Sabouni et al (24). The eta and etb loci can be detected from S. aureus isolated from children with skin diseases including SSSS, impetigo, or blisters (25). Notwithstanding the advancement of antibiotic therapy, SSSS display a significant mortality rate mainly among infants with secondary infections followed by epidermal loss and among adults with underlying diseases (26).

The *etd* gene is located within a 9.0-kb pathogenicity island (chromosomal site encoding virulence-associated factors), has also been found in this study, but was less common (20% of isolates) than other two exfoliative toxins. ETD-producing *S. aureus* are principally isolated from with suppurative skin infection such as cutaneous abscesses or furuncles, and not from SSSS (27). Recently, Mohseni et al. reported a frequency of 54% *S. aureus* harborin etd in Iran

Table 1.	The sequences,	product length	, annealing	temperature	(Tm)	of the
oligonucleotide	e primers used in	the study.				

Gene target	Sequences (5'-3')	Product (bp)	Tm (°C)	Reference
hla	CTGATTACTATCCAAGAAATTCGAT	210	53	(36)
	CTTTCCAGCCTACTTTTTTATCAGT			
hlb	GTGCACTTACTGACAATAGTGC	310	53	(36)
	GTTGATGAGTAGCTACCTTCAGT			
tsst-1	TTATCGTAAGCCCTTTTGTTG	398	46	(21)
	TAAAGGTAGTTCTATTGGAGTAGG			
eta	TTTGCTTTCTTGATTTGGATTC	464	51	(37)
	GATGTGTTCGGTTTGATTGAC			
etb	ACGGCTATATACATTCAATT	226	51	(37)
	TCCATCGATAATATACCTAA			
etd	GGGGAGACTATAGCTTCTGGTGTATTA	477	55.5	(37)
	TCCAACATGAATACCAACTAACTCT			

Table 2.The distribution of exotoxin genes among MRSA isolates from pediatricpatients.

No. Percent	virulence genes					
	hla	hlb	eta	etb	etd	tsst-1
10.25%	+	-	+	+	+	-
2.56%	+	+	+	-	-	+
7.69%	+	+	+	-	-	-
2.56%	-	-	-	+	-	+
5.12%	+	-	+	-	-	+
2.56%	+	+	+	-	+	-
10.25%	+	-	+	+	-	-
2.56%	+	+	+	-	+	-
2.56%	+	-	-	-	-	-
2.56%	+	-	-	-	+	+
2.56%	+	-	+	+	-	+
2.56%	+	-	+	-	+	+
2.56%	-	-	+	-	-	+
2.56%	+	+	+	+	-	+
20.51	+	-	+	-	-	-
10.25%	-	-	+	-	-	-
5.12%	+	-	+	-	+	-
2.56%	-	-	-	-	-	-
2.56%	+	+	-	-	-	+

(28). In a study conducted by Abimanyu et al. in India it was reported that none of the S. aureus isolates was found to carry *etd* gene but in our study all of our pediatrics patients had *etd* gene (29).

In our study we detected that 25.64% of the MRSA isolates were tsst-1 positive. Similarly, in the study conducted by Arabestani it was reported that tsst-1 was found in 30.61% of the *S. aureus* isolates (30). However, Esmaeili Benvidi et al. in 2017 reported that *tsst-1* was detected in 17% of the staphylococcal isolates from pediatric

patients in Tehran. In addition, Sina et al. previously described that *tsst*-1 was detected at a very low rate (1%) of isolated *S. aureus* in Iran (31). The disorders mediated by *tsst*-1, toxic shock syndrome, is a superantigen-mediated illness and characterized by rash, hypotension and multi-organ dysfunction (32). Infants and young children have a greater risk of developing severe toxic shock syndrome than adults. It is believed that this risk is associated with colonization by TSST-1-producing *S. aureus* in this population with insufficient antibody titers (33). If this illness is not cured quickly in children, the mortality rate can be high (34, 35). It is imported to find spread of MRSA strains and the virulence determinants among adults and pediatrics population but in Izzard Aglua et al. in 2018 reported just detection of MRSA strains and antibiotics patterns (36).

Conclusion

To the best of our knowledge, the current study is one of the few researches investigating the frequency of exotoxin genes of MRSA isolated from patients in Iran. In the present study, the results showed that the virulence determinants content of MRSA isolated from patients varies widely. Consequently, accurate and powerful epidemiological typing approaches such as pulsed-field gel electrophoresis (PFGE) and sequence typing multilocus (MLST) are recommended for active surveillance of MRSA strains and limiting the spread of epidemic clones within and between hospitals and to community settings. In addition, to avoid the spread of MRSA strains in community and healthcare settings, the decision should be made in conjunction with the infection control staff of the facility and must be clearly documented.

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Ethics approval and consent to participate

The study was conducted under the ethics approval code IR.SBMU.MSP.REC.1397.631.

Conflict of interest

No conflict of interests is declared.

References

- Tong SYC, Davis JS, Eichenberger E, et al. *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clin Microbiol Rev* 2015; 28(3):603-61.
- Bouvet C, Gjoni S, Zenelaj B, et al. *Staphylococcus aureus* soft tissue infection may increase the risk of subsequent staphylococcal soft tissue infections. *Int J Infect Dis* 2017; 60:44-8.
- Tong SY, Davis JS, Eichenberger E, et al. *Staphylococcus aureus* infections: epidemiology, pathophysiology, clinical manifestations, and management. *Clin Microbiol Rev* 2015; 28(3):603-61.
- Kaplan SL. Staphylococcus aureus infections in children: the implications of changing trends. Pediatrics. 2016; 137(4):e20160101.
- Oogai Y, Matsuo M, Hashimoto M, et al. Expression of virulence factors by *Staphylococcus aureus* grown in serum. *Applied Environment Microbiol* 2011; 77(22):8097-105.
- 6. Thomer L, Schneewind O, Missiakas D. Pathogenesis of *Staphylococcus aureus*

- 7. Naber CK. *Staphylococcus aureus* bacteremia: epidemiology, pathophysiology, and management *strategies*. *Clin Infect Dis* : an official publication of the Infectious Diseases Society of America 2009;48 Suppl 4:S231-7.
- Ferry T, Perpoint T, Vandenesch F, et al. Virulence determinants in *Staphylococcus aureus* and their involvement in clinical syndromes. *Current Infect Dis Reports* 2005; 7(6):420-8.
- Tappe D, Schulze MH, Oesterlein A, et al. Panton-Valentine leukocidin-positive Staphylococcus aureus infections in returning travelers. Am J Tropical Medicine Hygiene 2010; 83(4):748-50.
- Diep BA, Palazzolo-Ballance AM, Tattevin P, et al. Contribution of Panton-Valentine leukocidin in community-associated methicillin-resistant *Staphylococcus aureus* pathogenesis. *PloS one* 2008; **3**(9):e3198.
- Kobayashi SD, Malachowa N, DeLeo FR. Pathogenesis of *Staphylococcus aureus* abscesses. *Am J Pathol* 2015; **185**(6):1518-27.
- Dantes R, Mu Y, Belflower R, et al. National burden of invasive methicillin-resistant *Staphylococcus aureus* infections, United States, 2011. JAMA Intern Med 2013; 173(21):1970-8.
- Elhassan MM, Ozbak HA, Hemeg HA, et al. Absence of the *mecA* gene in methicillin resistant *Staphylococcus aureus* isolated from different clinical specimens in Shendi city, Sudan. *BioMed Res Inter* 2015; 2015:895860-.
- Mahdiyoun SM, Kazemian H, Ahanjan M, et al. Frequency of aminoglycoside-resistance genes in methicillin-resistant *Staphylococcus aureus* (MRSA) isolates from hospitalized patients. *Jundishapur J Microbiol* 2016; 9(8):e35052.

- Schmitz FJ, Fluit AC, Gondolf M, et al. The prevalence of aminoglycoside resistance and corresponding resistance genes in clinical isolates of staphylococci from 19 European hospitals. *J Antimicrobial Chemother* 1999; 43(2):253-9.
- 16. Houri H, Kazemian H, Sedigh Ebrahim-Saraie H, et al. Linezolid activity against clinical Gram-positive cocci with advanced antimicrobial drug resistance in Iran. *J Glob Antimicrob Resist* 2017; **10**:200-3.
- 17. Sina H, Ahoyo TA, Moussaoui W, et al. Variability of antibiotic susceptibility and toxin production of *Staphylococcus aureus* strains isolated from skin, soft tissue, and bone related infections. *BMC Microbiol* 2013; **13**:188.
- 18. Otto M. Staphylococcus aureus toxins. Current Opinion Microbiol 2014; 17:32-7.
- 19. Argudin MA, Mendoza MC, Vazquez F, et al. Exotoxin gene backgrounds in bloodstream and wound *Staphylococcus aureus* isolates from geriatric patients attending a long-term care Spanish hospital. *J Med Microbiol* 2011; **60**(Pt 11):1605-12.
- 20. Francis JS, Doherty MC, Lopatin U, et al. Severe community-onset pneumonia in healthy adults caused by methicillin-resistant *Staphylococcus aureus* carrying the Panton-Valentine leukocidin genes. *Clin Infect Dis*: an official publication of the Infectious Diseases Society of America 2005; **40**(1):100-7.
- Benvidi ME, Houri H, Ghalavand Z, et al. Toxin production and drug resistance profiles of pediatric methicillin-resistant *Staphylococcus aureus* isolates in Tehran. J Infect Develop Countries 2017; 11(10):759-65.
- 22. Emaneini M, Beigverdi R, van Leeuwen WB, et al. Prevalence of Methicillin-Resistant *Staphylococcus aureus* isolated from burn patients: A systematic review and meta-analysis. *J Glob Antimicrob Resist*

2017; **12**:202-6.

- Mahdiyoun SM, Kazemian H, Ahanjan M, et al. Frequency of aminoglycoside-resistance genes in methicillin-resistant *Staphylococcus aureus* (MRSA) isolates from hospitalized patients. *Jundishapur J Microbiol* 2016; 9(8):e35052.
- 24. Sabouni F, Mahmoudi S, Bahador A, et al. Virulence factors of *Staphylococcus aureus* isolates in an Iranian referral children's hospital. *Osong Public Health Res Perspectives* 2014; 5(2):96-100.
- 25. Mishra AK, Yadav P, Mishra A. A systemic review on staphylococcal scalded skin syndrome (SSSS): A rare and critical disease of neonates. *Open Microbiol J* 2016; **10**:150-9.
- Jeyakumari D, Gopal R, Eswaran M, et al. Staphylococcal scalded skin syndrome in a newborn. J Global Infect Dis 2009; 1(1):45-7.
- 27. Yamasaki O, Tristan A, Yamaguchi T, et al. Distribution of the exfoliative toxin D gene in clinical *Staphylococcus aureus* isolates in France. *Clin Microbiol Infect*: the official publication of the European Society of Clinical Microbiology and Infectious Diseases 2006; **12**(6):585-8.
- 28. Mohseni M, Rafiei F, Ghaemi EA. High frequency of exfoliative toxin genes among *Staphylococcus aureus* isolated from clinical specimens in the north of Iran: Alarm for the health of individuals under risk. *Iran J Microbiol* 2018; **10**(3):158-65.
- 29. Abimanyu N, Murugesan S, Krishnan P. High prevalence of exfoliative toxins among carrier isolates of *Staphylococcus aureus* from healthy individuals from various communities in Chennai, south India. *Indian J Microbiol* 2013; **53**(3):288-90.
- 30. Arabestani MR, Kazemian H, Tabar ZK, et al. Prevalence of virulence genes, *agr* and antimicrobial resistance of *Staphylococcus aureus* isolated from food and dairy products

in Hamadan, Iran. *Der Pharmacia Lettre* 2016; **8**(8):62-7.

- 31. Sina H, Ahoyo TA, Moussaoui W, et al. Variability of antibiotic susceptibility and toxin production of *Staphylococcus aureus* strains isolated from skin, soft tissue, and bone related infections. *BMC Microbiol* 2013; **13**:188.
- 32. Strom MA, Hsu DY, Silverberg JI. Prevalence, comorbidities and mortality of toxic shock syndrome in children and adults in the USA. *Microbiol Immunol* 2017; 61(11):463-73.
- 33. Quan L, Morita R, Kawakami S. Toxic shock syndrome toxin-1 (TSST-1) antibody levels in Japanese children. Burns : Journal of the International Society for Burn Injuries 2010; 36(5):716-21.
- Roy P, Sahni AK, Kumar A. A fatal case of staphylococcal toxic shock syndrome. *Med J, Armed Forces India* 2015; 71(Suppl 1):S107-S10.
- 35. Aglua I, Jaworski J, Drekore J, et al. Methicillin resistant *Staphylococcus aureus* in Melanesian children with haematogenous osteomyelitis from the central highlands of Papua New Guinea. *Int J Pediatr* 2018; 6(10):8361-70.
- 36. Yu F, Liu Y, Lv J, et al. Antimicrobial susceptibility, virulence determinant carriage and molecular characteristics of *Staphylococcus aureus* isolates associated with skin and soft tissue infections. *Brazilian J Infect Dis* 2015; **19**(6):614-22.
- Futagawa-Saito K, Makino S, Sunaga F, et al. Identification of first exfoliative toxin in *Staphylococcus pseudintermedius. FEMS Microbiol letters* 2009; 301(2):176-80.